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HELLER EHRMAN WHITE & MCAULIFFE LLP
4350 LA JOLLA VILLAGE DRIVE
7TH FLOOR
SAN DIEGO, CA 92122-1246

EXAMINER

GOLDBERG, JEANINE ANNE

ART UNIT PAPER NUMBER

1634

DATE MAILED: 11/06/2002

19

Please find below and/or attached an Office communication concerning this application or proceeding.

Office Action Summary

Application No.

09/601,645

Applicant(s)

DAHM ET AL.

Examiner

Jeanine A Goldberg

Art Unit

1634

-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --
Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) FROM
THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If the period for reply specified above is less than thirty (30) days, a reply within the statutory minimum of thirty (30) days will be considered timely.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133).
- Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

- 1) ☒ Responsive to communication(s) filed on 1/9/02; 2/25/02.
- 2a) ☐ This action is **FINAL**. 2b) ☒ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

- 4) ☒ Claim(s) 1-70 is/are pending in the application.
- 4a) Of the above claim(s) 39-51 and 68 is/are withdrawn from consideration.
- 5) ☐ Claim(s) _____ is/are allowed.
- 6) ☒ Claim(s) 1-38, 52-67, 69-70 is/are rejected.
- 7) ☐ Claim(s) _____ is/are objected to.
- 8) ☐ Claim(s) _____ are subject to restriction and/or election requirement.

Application Papers

- 9) ☐ The specification is objected to by the Examiner.
- 10) ☐ The drawing(s) filed on _____ is/are: a) ☐ accepted or b) ☐ objected to by the Examiner.
- Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
- 11) ☐ The proposed drawing correction filed on _____ is: a) ☐ approved b) ☐ disapproved by the Examiner.
- If approved, corrected drawings are required in reply to this Office action.
- 12) ☐ The oath or declaration is objected to by the Examiner.

Priority under 35 U.S.C. §§ 119 and 120

- 13) ☒ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☒ All b) ☐ Some * c) ☐ None of:
1. ☐ Certified copies of the priority documents have been received.
2. ☐ Certified copies of the priority documents have been received in Application No. _____.
3. ☒ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).
- * See the attached detailed Office action for a list of the certified copies not received.
- 14) ☐ Acknowledgment is made of a claim for domestic priority under 35 U.S.C. § 119(e) (to a provisional application).
- a) ☐ The translation of the foreign language provisional application has been received.
- 15) ☐ Acknowledgment is made of a claim for domestic priority under 35 U.S.C. §§ 120 and/or 121.

Attachment(s)

- 1) ☒ Notice of References Cited (PTO-892)
- 2) ☐ Notice of Draftsperson's Patent Drawing Review (PTO-948)
- 3) ☐ Information Disclosure Statement(s) (PTO-1449) Paper No(s) _____.
- 4) ☐ Interview Summary (PTO-413) Paper No(s). _____.
- 5) ☐ Notice of Informal Patent Application (PTO-152)
- 6) ☐ Other: _____.

DETAILED ACTION

1. This action is in response to the papers filed February 25, 2002; January 9, 2002. Currently, claims 1-70 are pending. Claims 39-51, 68 have been withdrawn as drawn to non-elected subject matter.
2. Any objections and rejections not reiterated below are hereby withdrawn.
3. Applicant's arguments with respect to the previous rejections are moot in view of the new grounds of rejection.

Election/Restrictions

4. As provided in the decision, mailed April 16, 2002, of the petition filed December 26, 2002, Claims 1-38, 52-67, 69-70 are under examination. Thus an action on the merits follows.

Claim Rejections - 35 USC § 102

The following is a quotation of the appropriate paragraphs of 35 U.S.C. 102 that form the basis for the rejections under this section made in this Office action:

A person shall be entitled to a patent unless –

(a) the invention was known or used by others in this country, or patented or described in a printed publication in this or a foreign country, before the invention thereof by the applicant for a patent.

(b) the invention was patented or described in a printed publication in this or a foreign country or in public use or on sale in this country, more than one year prior to the date of application for patent in the United States.

(e) the invention was described in a patent granted on an application for patent by another filed in the United States before the invention thereof by the applicant for patent, or on an international application by another who has fulfilled the requirements of paragraphs (1), (2), and (4) of section 371(c) of this title before the invention thereof by the applicant for patent.

The changes made to 35 U.S.C. 102(e) by the American Inventors Protection Act of 1999 (AIPA) do not apply to the examination of this application as the application being examined was not (1) filed on or after November 29, 2000, or (2) voluntarily published under 35 U.S.C. 122(b). Therefore, this application is examined under 35 U.S.C. 102(e) prior to the amendment by the AIPA (pre-AIPA 35 U.S.C. 102(e)).

5. Claims 1-2, 4, 7-11, 14-17, 37-38, 52-56, 67 are rejected under 35 U.S.C. 102(e) as being anticipated by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997).

It is noted that only the specification for the patent has been provided because the sequences are not relied upon in the rejection and are very extensive in length.

Cech et al. (herein referred to as Cech) teaches methods of quantitating tumor cells in a body fluid by concentrating tumor cells in a sample of body fluid, amplifying mRNA coding for the catalytic subunit of telomerase and quantitatively determining the amount of amplified nucleic acid. Specifically, Cech teaches methods of diagnosing cancer in a patient by obtaining a biological sample from the patient and detecting a hTERT gene product in the patient sample, where the detection of the hTERT gene product in the sample is correlated with a diagnosis of cancer (col. 6, lines 20-40). Cech also teaches that the determination of an hTERT gene, mRNA or protein level above normal or standard range is indicative of the presence of telomerase-positive cells, or immortal of which certain tumor cells are examples (col. 99, lines 5-20). Cech specifically teaches that hTERT gene or gene product (i.e., mRNA or polypeptide) is preferably detected and/or quantified in a biological sample (col. 104, lines 59-65).

Art Unit: 1634

Cech teaches that biological samples include blood, blood cells, body fluids, e.g., urine, sputum, amniotic fluid, blood, peritoneal fluid, pleural fluid, semen (col. 104, lines 65-68)(limitations of Claim 11, 16-17). Cech teaches that cells or tissues may be fractionated before analysis, for example by a cell sorter may be used to sort cells according to characteristics such as expression of a surface antigen (col. 105, lines 10-12). Cech teaches that nucleic acids may be isolated from the cell by any separation of the species or target to be detected from any other substance in the mixture (col. 105, lines 55-65)(limitations of Claim 4). Cech teaches that the assay for detection of hTERT are amplification based to amplify all or part of an hTERT gene or transcript where the amplification product is then detected directly or indirectly (col. 106, lines 45-55). Cech teaches primers useful for PCR amplification of hTERT are provided in Table 2 (col. 107, lines 1-5). Cech teaches that amplified products may be directly analyzed by size (gel electrophoresis); by hybridization to a target nucleic acid immobilized on a solid support; by sequencing; by detection of a fluorescent, phosphorescent, or radioactive signal (col. 108, lines 1-5)(limitations of Claim 8-9, 55-56). Cech teaches that in one possible embodiment PCR amplification is carried out in a 50ul solution containing the nucleic acid sample (e.g., cDNA obtained through reverse transcription of hTERT RNA), dNTP, hTERT specific PCR primers, Taq polymerase, PCR buffer (col. 107, lines 12-25)(limitations of Claim 2, 14, 15). Cech teaches that quantification methods may include the co-amplification reactions to allow for normalization of the cell number in a sample as compared to the amount of hTERT in the sample (col. 108, lines 45-65)(limitations of Claim 7, 10, 54).

Claim Rejections - 35 USC § 103

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negated by the manner in which the invention was made.

6. This application currently names joint inventors. In considering patentability of the claims under 35 U.S.C. 103(a), the examiner presumes that the subject matter of the various claims was commonly owned at the time any inventions covered therein were made absent any evidence to the contrary. Applicant is advised of the obligation under 37 CFR 1.56 to point out the inventor and invention dates of each claim that was not commonly owned at the time a later invention was made in order for the examiner to consider the applicability of 35 U.S.C. 103(c) and potential 35 U.S.C. 102(e), (f) or (g) prior art under 35 U.S.C. 103(a).

7. Claims 18-28, 34-36, 60-64, 69-70 are rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997) in view of Van Vlasselaer et al (US Pat. 5,648,223, July 1997).

It is noted that only the specification for the patent has been provided because the sequences are not relied upon in the rejection and are very extensive in length.

Cech et al. (herein referred to as Cech) teaches methods of quantitating tumor cells in a body fluid by concentrating tumor cells in a sample of body fluid, amplifying

mRNA coding for the catalytic subunit of telomerase and quantitatively determining the amount of amplified nucleic acid. Specifically, Cech teaches methods of diagnosing cancer in a patient by obtaining a biological sample from the patient and detecting a hTERT gene product in the patient sample, where the detection of the hTERT gene product in the sample is correlated with a diagnosis of cancer (col. 6, lines 20-40). Cech also teaches that the determination of an hTERT gene, mRNA or protein level above normal or standard range is indicative of the presence of telomerase-positive cells, or immortal of which certain tumor cells are examples (col. 99, lines 5-20). Cech specifically teaches that hTERT gene or gene product (i.e., mRNA or polypeptide) is preferably detected and/or quantified in a biological sample (col. 104, lines 59-65). Cech teaches that biological samples include blood, blood cells, body fluids, e.g., urine, sputum, amniotic fluid, blood, peritoneal fluid, pleural fluid, semen (col. 104, lines 65-68)(limitations of Claim 11, 24-28). Cech teaches that cells or tissues may be fractionated before analysis, for example by a cell sorter may be used to sort cells according to characteristics such as expression of a surface antigen (col. 105, lines 10-12). Cech teaches that nucleic acids may be isolated from the cell by any separation of the species or target to be detected from any other substance in the mixture (col. 105, lines 55-65)(limitations of Claim 4). Cech teaches that the assay for detection of hTERT are amplification based to amplify all or part of an hTERT gene or transcript where the amplification product is then detected directly or indirectly (col. 106, lines 45-55). Cech teaches primers useful for PCR amplification of hTERT are provided in Table 2 (col. 107, lines 1-5). Cech teaches that amplified products may be directly analyzed by size (gel

electrophoresis); by hybridization to a target nucleic acid immobilized on a solid support; by sequencing; by detection of a fluorescent, phosphorescent, or radioactive signal (col. 108, lines 1-5)(limitations of Claim 8-9). Cech teaches that in one possible embodiment PCR amplification is carried out in a 50ul solution containing the nucleic acid sample (e.g., cDNA obtained through reverse transcription of hTERT RNA), dNTP, hTERT specific PCR primers, Taq polymerase, PCR buffer (col. 107, lines 12-25)(limitations of Claim 2, 14, 15). Cech teaches that quantification methods may include the co-amplification reactions to allow for normalization of the cell number in a sample as compared to the amount of hTERT in the sample (col. 108, lines 45-65)(limitations of Claim 7, 10).

Cech does not specifically teach enrichment by centrifugation to collect tumor cells.

However, Van Vlasselaer teaches methods for enriching tumor cells prior to analyzing. Percoll and Ficoll were routinely used in the art as cell separation media (col. 4, lines 60- col. 5, lines 5)(limitations of claim 23). Van Vlassalaer teaches that the medium density is adjusted to the density of the cell type (col. 9, lines 47-66, col. 14, Example 6.1.1 and 6.1.2)(limitations of Claim 21, 22, 24 and 62-64). Van Vlassalaer teaches that a large volume of complete blood may be directly placed on the gensity gradient. Peripheral blood may be collected in anti-coagulant containing tubes (col. 4, lines 37-40)(limitations of Claim 24-27). Van Vlassalaer teaches that for breast tumor cells, the specific density was adjusted within 0.0005 g/ml of the specific density of the tumor cells and the centrifugation speed is at a gravitational force sufficient to pellet the cells (col. 16, lines 61-67). Additionally, Van Vlasselaer teaches methods that may

cause cells to be heavier than their normal density so that they are pelleted during centrifugation, namely linking a heavy particle such as a binding agent to selected cells. Van Vlasselaer teaches that centrifugation is carried out in a tube divided by a barrier wherein the barrier is an annular ring (col. 5, lines 30-54).

Therefore, it would have been prima facie obvious to one of ordinary skill in the art to have modified the method of Cech with the enrichment method of Van Vlasselaer prior to molecular analysis. Van Vlasselaer teaches that after centrifugation is performed and cells are collected, tumor cell may be screened by molecular means (col. 4 lines 20-22). Therefore, the ordinary artisan would have been motivated to have performed the rapid and high yield isolation or enrichment of tumor cells prior to analysis by the method of Cech for the increased sensitivity and efficiency. The numerous means of enrichment and isolation of Van Vlasselaer would be obvious means to enriching cells in a blood sample. Therefore, it would have been obvious to the ordinary artisan to employ a cell separation medium that was proven in the art and readily available.

With regard to Claims 21, 22, 24, 62-64, it would have been obvious to the ordinary artisan to adjust the density of the cell separation medium and centrifugation speed according to the type of tumor cell to be concentrated. Van Vlasselaer specifically teaches that methods for determining the specific density of a given tumor cell is described infra (col. 4, lines 10-15). Additionally, Van Vlasselaer teaches methods that may cause cells to be heavier than their normal density so that they are pelleted during centrifugation, namely linking a heavy particle such as a binding agent to

selected cells. Therefore, the determination of cell densities requires routine experimentation. With regard to the densities of certain cells, these are routinely optimizable based upon the desired parameters, since Van Vlasselaer teaches how densities may be determined the optimization of the workable density is not inventive. As noted in *In re Aller*, 105 USPQ 233 at 235, "More particularly, where the general conditions of a claim are disclosed in the prior art, it is not inventive to discover the optimum or workable ranges by routine experimentation." It would have been obvious to provide a substance that prevents platelets from sticking to the tumor cells and to remove the platelets as routinely practiced in the art.

8. Claim 3 is rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997) above in view of Gwynn et al. (US Pat. 6,025,156, February 2000).

Cech does not specifically teach using DNAase for removal of DNA from a sample.

However, Gwynn et al. (herein referred to as Gwynn) teaches using DNAase for removal of DNA from RNA samples. Once the DNAase was added and DNA was removed, RNA was pelleted and reverse transcribed into cDNA (col. 31, lines 50-65).

Therefore, it would have been prima facie obvious to one of ordinary skill in the art to have modified the method of generating cDNA of Cech with the teachings of Gwynn. Gwynn teaches addition of DNAase facilitates the removal of DNA from the sample such that RNA may be obtained. Therefore, the ordinary artisan would have

realized that in order to remove DNA from RNA samples so that the RNA may be in turn transcribed, DNAase may be added.

9. Claims 29 is rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997) above in view of Shelby (GB 2 260 811, April 1993).

Shelby teaches that diagnosis or monitoring of cancer or a malignant tumor may be effected by the detection of mRNA in a sample such as peripheral blood where the cells are not normally present and testing the sample (abstract). The detection technique of Shelby involves extracting the total cellular mRNA in a sample using reverse transcriptase to prepare cDNA, then carrying out PCR with appropriate primers so as to selectively amplify the cDNA (abstract). Shelby teaches cooling after centrifugation was routinely practice in the art (page 10, last paragraph).

Therefore, it would have been prima facie obvious to one of ordinary skill in the art to have modified the method of centrifugation by the allowing of the sample to cool following centrifugation.

10. Claims 5-6 are rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997) above in view of Gelmini et al. (Clinical Chemistry, Vol. 43, No. 5, pages 752-758, 1997).

Cech does not specifically teach the continuous monitoring of PCR reaction.

However, Gelmini teaches methods of quantitative polymerase chain reaction which is quantitative, accurate, and time-saving. The method of Gelmini uses fluorogenic probes to assess amplification. Gelmini teaches that during PCR cycling, the probe specifically hybridizes to the corresponding template and then is cleaved and results in increase of fluorescence emission of the reporter dye. The increased of fluorescence is proportional to the amount of the specific PCR product (page 753, col. 2). Gelmini teaches that the fluorescence was measured with a luminescence spectrometer (page 754, col. 1).

Therefore, it would have been prima facie obvious to one of ordinary skill in the art to have modified the method of Cech for amplification with the method of Gelmini for the real time quantitative PCR. The ordinary artisan would have been motivated to have applied the method of Gelmini because Gelmini teaches the TaqMan PCR assay gave accurate, quantitative results.

11. Claims 13 are rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997).

Cech does not specifically teach SEQ ID NO: 1 and 2 however, Cech teaches numerous primers suitable for PCR amplification of hTERT.

The nucleic acid sequences of SEQ ID NO: 1 and 2 are functional equivalents to the primers taught by Cech. The ordinary artisan would have recognized that primer designed to amplify all or part of an hTERT gene may be used. Cech teaches the parameters needed to design appropriate primers for hTERT. For example, Cech

teaches that the primer are sufficiently complementary to the hTRT gene. The primers are typically at least 6 bases in length, typically between about 12 and about 50 bases (col. 106, lines 45-68). Cech teaches that one of skill in the art having the disclosure will be able, using routine methods will select primer to amplify all or any portion of hTRT gene. Therefore, SEQ ID NO: 1 and 2 of the instant application were merely selected by the routine methods provided by Cech for the amplification of all or part of the hTRT nucleic acid.

12. Claims 12, 57-59 are rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997) above in view of Melvin et al. (WO 97/12246, April 1997).

Cech does not specifically teach using controls.

However, Melvin et al. (herein referred to as Melvin) teaches that in RT-PCR experiments B-actin was used as a positive control. Melvin teaches that the negative control was sterile water in place of cDNA (page 17).

Therefore, it would have been prima facie obvious to one of ordinary skill in the art to have modified the method of Cech to include controls as taught by Melvin. Controls are essential in each scientific experiment to ensure that the results obtained are due to the experiment and not due to external factors. Therefore, the ordinary artisan would be motivated to have used controls in the study of Cech.

13. Claims 30-33, 65-66 are rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997) in view of Van Vlasselaer et al (US Pat. 5,648,223, July 1997) as applied to Claims 18-28, 60-64 above, and further in view of Oka et al (US Pat. 5,298,165, March 1994).

Neither Cech nor Van Vlasselaer specifically teach centrifugation with filters of porous barriers which have certain properties.

However Oka et al. (herein referred to as Oka) teaches that filtration of blood may be effected with different membranes, filters or porous barriers. Oka teaches that the average pore size of one of the filters is preferably from 4 to 25 um (col. 10, lines 40-45). Additionally, Oka teaches numerous different filters with different thickness including thickness of 2 mm, 5 mm.

Therefore, it would have been prima facie obvious to one of ordinary skill in the art to have modified the method of Cech in view of Van Vlasselaer to enrich using a porous barrier, filter or sieve as taught by Oka for the express benefit of enriching or isolating cells. The filters of Oka are representative of filters taught in the art. As exemplified by Oka the specific specifications of the filter are dependent upon the material wishing to be isolated. Therefore, with regard to the pore size and thickness of filters, these are routinely optimizable based upon the desired parameters, since Oka teaches how densities may be determined the optimization of the workable pore size and thickness is not inventive. As noted in *In re Aller*, 105 USPQ 233 at 235, "More particularly, where the general conditions of a claim are disclosed in the prior art, it is not inventive to discover the optimum or workable ranges by routine experimentation."

Art Unit: 1634

Conclusion

14. No claims allowable over the art.


15. Any inquiry concerning this communication or earlier communications from the examiner should be directed to examiner Jeanine Goldberg whose telephone number is (703) 306-5817. The examiner can normally be reached Monday-Friday from 8:00 a.m. to 5:30 p.m.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Gary Jones, can be reached on (703) 308-1152. The fax number for this Group is (703) 305- 3014.

Any inquiry of formal matters can be directed to the patent analyst, Pauline Farrier, whose telephone number is (703) 305-3550.

Any inquiry of a general nature should be directed to the Group receptionist whose telephone number is (703) 308-0196.

Jeanine Goldberg
November 4, 2002


W. Gary Jones
Supervisory Patent Examiner
Technology Center 1600